



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

There's plenty of room at the bottom: Low radiation as a biological extreme

Citation for published version:

Wadsworth, J, Cockell, CS, Murphy, ASJ, Nilima, A, Paling, SM, Meehan, E, Toth, C, Scovell, P & Cascorbi, L 2020, 'There's plenty of room at the bottom: Low radiation as a biological extreme', *Frontiers in Astronomy and Space Sciences*. <https://doi.org/10.3389/fspas.2020.00050>

Digital Object Identifier (DOI):

[10.3389/fspas.2020.00050](https://doi.org/10.3389/fspas.2020.00050)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Peer reviewed version

Published In:

Frontiers in Astronomy and Space Sciences

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



There's plenty of room at the bottom: Low radiation as a biological extreme

Jennifer Wadsworth^{1*}, Charles S. Cockell¹, Alexander StJ Murphy¹, Athoy Nilima¹, Sean Paling², Emma Meehan², Christopher Toth², Paul Scovell², Leander Cascorbi³

¹School of Physics and Astronomy, University of Edinburgh, UK

²Boulby Underground Laboratory, Redcar & Cleveland, UK

³Trinity College, Oxford University, UK

*** Correspondence:**

Jennifer Wadsworth
jenn@wadsworth.ch

Abstract

The effects of high radiation as a biological extreme have historically been, and continue to be, extensively researched in the fields of radiation biology and astrobiology. However, the absence of radiation as an extreme has received relatively limited attention from the scientific community, with its effects on life remaining unclear. The currently accepted model of the radiation dose-damage relationship for organisms is the linear no-threshold (LNT) model, which predicts a positive linear correlation between dose and damage that intercepts at zero dose corresponding to zero damage. Despite its wide-spread implementation, the LNT model is continuously being challenged by various new models, with the hormesis model as one of its main competitors. This model also postulates damage at high doses but, in contrast to the LNT model, it predicts beneficial stimulation of growth at low doses. Experiments to date have not yet been able to conclusively validate or dismiss either of these models. The aim of the collaborative Subsurface Experiment of Life in Low Radiation (SELLR) project was to test these competing models on prokaryotes in a well-characterised environment and provide a robust experimental set up to investigate low radiation in terrestrial and non-terrestrial environments. Bacterial growth assays using *Bacillus subtilis* and *Escherichia coli* were performed under ultra low ionising radiation in the Boulby International Subsurface Astrobiology Laboratory (BISAL) facilities of the Boulby Underground Laboratory at Boulby mine (Redcar & Cleveland, UK) and were used to investigate effects on viability and signs of preconditioning. No significant effect on bacterial growth was observed from exposure to radiation doses ranging from 0.01 times the levels of background radiation typically found in terrestrial surface environments to 100 times that background. Additionally, no preconditioned susceptibility to stress was observed in the bacterial strains grown in sustained low radiation. These data suggest that the extremes of low radiation do not alter growth parameters of these two organisms and that an improved model should be considered for prokaryotes, consisting of a dose-damage response with a threshold at ultra low radiation. We discuss the implications of these data for low radiation as a novel microbiological “extreme”.

1. Introduction

Radiation biology research is mostly conducted in the context of effects of, and protection from, high radiation exposure such as from radioactive fallout and medical treatments (e.g. Finch et al., 2007; de Gonzalez et al., 2004; Preston et al., 2004). In this regard, the field of Astrobiology is no exception with a seemingly exclusive focus on the effects of the high radiation conditions of low Earth orbit, interplanetary space and other planetary environments on organisms (e.g. Horneck, 1992; Moeller et al., 2017; de Vera et al., 2019). The historical and continuing scientific focus on high radiation is by no means unfounded, as the connection between irradiation from an ionising or an energetic radiation source and biological damage, resulting in mutations or cell death, is well established (Lorenz, 1944; Wanebo et al., 1968; Chang et al., 1985; Ward, 1988). However, extreme opposites of physical variables exist and, to use the title of Richard Feynman's 1959 address to the annual American Physical Society meeting at Caltech (Feynman, 1960), there's "plenty of room at the bottom" of the radiation dose range to explore as a physical extreme. In order to build a holistic understanding of how environmental parameters shape the viability potential of life on Earth and, by extension, on other planetary bodies, all parameter extremes should be examined.

All life on Earth is constantly being subjected to a low dose of naturally occurring radiation in various forms. For example, non-ionising ultraviolet (UV) light (UVA = 315 - 400 nm, UVB = 280 - 315 nm) from the sun can pass through the Earth's atmosphere and reach us, sometimes causing skin melanomas (International Agency for Research on Cancer, 1992). Of the naturally occurring radiation, ionising radiation is of particular significance due to its increased potential for causing biological damage (e.g. Prise et al., 2001). Rock radionuclides from various long-lived radioisotopes, such as uranium, thorium and potassium, and cosmic rays are sources of natural ionising radiation (Hamilton, 1989; Shahbazi-Gahrouei et al., 2013). Moreover, airborne radon and its decay radionuclides contribute to more than half the total average worldwide dose rate of naturally occurring radiation of ~ 2.8 mGy/y. With additional man-made radiation, the mean global surface radiation dose rate is estimated to be ~ 3.01 mGy/y (United Nations, 2008), although this can vary locally e.g. 0.6 mGy/y measured at the Boulby mine surface to 260 mGy/y in Ramsar, Iran (Ghiassi-Nejad et al., 2002). Usually, doses/dose rates below the global or local background dose average are referred to as "low" or "ultra low" doses in low radiation studies.

There are currently three main ideas on how a low radiation environment might influence an organism's viability: 1) Any amount of radiation, however small, will negatively impact viability and continue to do so in a linear manner ("linear no-threshold" model), 2) A hyperbolic dose-damage response to low radiation in the form of either hypersensitivity or positive stimulation (the latter being the "hormesis" model), which would be a unique property among extreme conditions. In the hormesis model, low levels of ionising radiation are therefore beneficial for life, and 3) At some low level of radiation there is no effect on life, with a threshold below which there is no discernible effect on cellular physiology or growth parameters. However, above the determined threshold the radiation dose-damage continues in a linear manner. This is known as the "threshold" model (fig. 1) (e.g. Mancuso et al., 2012; Siasou et al., 2017). Since the mid-20th century, the generally accepted model for describing an organism's radiation dose-response relationship has been the linear no-threshold (LNT) model (Muller, 1946) (fig. 1), which was further supported by the National Research Council

(NRC, 2006). The LNT model is largely based on studies in above-background radiation conditions, i.e. high radiation exposure. Radiation-induced mutations in rats in the early 20th century (Olson and Lewis, 1928), and subsequent research on fruit fly exposure to high radiation doses (10^3 times background dose) served as the basis for development of the LNT model (Muller, 1954). However, subsequent studies have indicated that when dealing with lower doses, the LNT model may not be applicable (e.g. Calabrese and Baldwin 2003; Feinendegen et al., 2007; Smith, et al., 2011; Shamoun, 2016; Shibamoto, 2018; Costantini and Borremans, 2019).

Despite the long-standing application of the LNT model, it is strongly contested in the literature by proponents of the hormesis (hormetic) model (fig. 1), who argue that the LNT model is not representative of the cellular response to lower radiation doses and dose rates (e.g. Feinendegen et al., 2007). The principle of hormesis is based on a low dose stimulatory effect of a physical or chemical component that, at high doses, has an inhibitory effect (Stebbing, 1982; Lorenz, 1950; Mine et al., 1990). Additional, less prevalent hypotheses include the “threshold” and “sensitisation/hypersensitivity” models, with the former proposing a lack of impact below a certain dose range and the latter serving as an opposite to hormesis, implying an increased negative effect on viability at low dose exposure (fig. 1). Although it is reasonable to presume the LNT model is also applicable to prokaryotes at higher doses of radiation, there is still no definitive consensus within the scientific community on the effects of low radiation exposure on microbes (fig. 1).

Here we present data from the Subsurface Experiment of Life in Low Radiation (SELLR) project on the effects of below background radiation on the growth of *Bacillus subtilis* and *Escherichia coli*. We also describe a robust experimental set-up that allows for the study of prokaryotic biology at below background levels of radiation using an underground astrobiology laboratory. The aim of the Subsurface Experiment of Life in Low Radiation (SELLR) project, described in this work, is to provide a well-controlled and characterised experimental set up with minimal variables in which comparable bacterial growth experiments can be carried out. The robust set up establishes a baseline study for future work simulating low radiation (planetary) environments.

2. Materials and Methods

Previous low radiation experiments (i.e. below the local terrestrial surface radiation doses) have been carried out in various facilities by research groups examining a variety of cells and with different experimental set-ups. Furthermore, when establishing and working in a low radiation experimental set up there are additional factors to those of standard experiments to consider. This and the lack of a universal, gold-standard low radiation set up has continued to impede the comparability of data in low radiation literature. Table 1 illustrates some of the potential experimental difficulties in a low radiation set up and the experimental design adjustments that were adopted for the SELLR project to avoid the outlined issues.

2.1. Experimental overview

Two prokaryotic model organisms were grown in the low radiation, underground experimental set-up that consisted of two identical plate readers housed in a lead “castle” to further reduce radiation

exposure. Within the castle, the plate readers were separated by a lead brick wall, such that one half of the castle provided the ultra low radiation environment whilst the other half provided an artificially created background radiation environment mimicking that of the local surface, established using a ^{137}Cs source (fig. 2). The model organisms were then grown in the plate readers for varying lengths of time in the low radiation environment and the artificial background radiation environment, which served as the control. After growth in either radiation condition, the organisms were briefly exposed to UVC to test potential differences in stress response.

2.2. Location and radiation set-up

Unless specifically stated, all experiments were carried out in the Boulby Underground Laboratory at the Boulby Mine, Redcar & Cleveland UK. At 1.1 km depth, the rock accounts for a cosmic ray attenuation by a factor of 10^6 (Reichhart et al., 2013). Any changes caused by the attenuation to the cosmic ray energy spectrum are assumed to be irrelevant as the cosmic rays are of high enough energies to be minimally ionising, such that energy deposition by muons and secondary neutron spallation rates will scale linearly with absolute cosmic ray flux.

With the cosmic ray flux minimised, the Boulby Laboratory is particularly well-suited to provide radiation protection as the local salt has low natural background radiation, with low levels of γ -ray and neutron emission. Moreover, the salt and general geographical area of Boulby have a low rate of radon production, resulting in an airborne background level of 2.4 Bq/m^3 (Araújo et al., 2012). To establish an ultra low radiation environment, the experiments were additionally encased in a radon-free lead-lined container (**castle**). This resulted in a 100-fold reduction of the average local ionising surface radiation dose rate (see the following method sections for details on the radiation environment). To reduce systematic uncertainties, measurements were performed in two experimental stations, formed by partitioning the lead-lined container in to two equal halves, separated by a 10 cm-thick wall formed of low-radioactivity lead bricks. Identical SPECTROstar Nano plate readers (BMG Labtech) were then positioned in each half of the lead-lined box (fig. 2).

To achieve elevated (above low background radiation doses), a ^{137}Cs source was placed above one of the plate readers to artificially generate surface radiation doses, and up to 100 times higher, while with no such source, the samples in the other plate reader provided well-defined controls. The ^{137}Cs source was used to approximate ionising radiation biological damage on the surface, recognising that it is difficult to artificially reconstruct the complete spectrum of natural radiation of the surface in a confined subsurface laboratory environment. The temperature of both plate readers was internally controlled, the outside temperature was constant at 22°C ($\pm 1^\circ\text{C}$) and 82 % relative humidity (± 2 %).

2.3. Artificial background and low radiation environments

Theoretical dose rates:

Both the low dose and control samples were located in the same sub-surface experimental set up (described in section 2.2) to avoid differences in environmental conditions.

Control samples were exposed to a 15.5 MBq ($\pm 0.1 \text{ MBq}$) ^{137}Cs γ -source to approximate a natural surface background radiation dose rate. The collimated but unattenuated source provided a dose rate

There's plenty of room at the bottom: Low radiation as a biological extreme

of $D = 27.8 \text{ mGy/y}$ ($\pm 0.506 \text{ mGy/y}$) at a 59 cm height from the top of the 96-well plate, which was lowered to 1 mGy/y by placing lead sheets between the source and samples. It was calculated that 11 lead sheets (3.3 cm total thickness) would be required to attenuate radiation from the ^{137}Cs γ -source to obtain a final dose rate comparable to that measured at the local surface (approx. 0.18 - 0.6 mGy/y).

To generate higher dose rates $D(r)$ of $10 \times$ and $100 \times$ background, the inverse square rule is applied.

$$D(r) = \left(\frac{59 \text{ cm}}{r} \right)^2 \times D(59 \text{ cm})$$

As the distance r between source and sample is decreased, the difference in local dose rates across the plate becomes more significant, i.e. as the source is moved closer to the samples there is a larger discrepancy between the dose rate of the centre of the plate compared to that of the periphery. It was calculated that for our maximum dose rate used (100 mGy/y) the maximum difference in dose rate across the plate would be 3.3 mGy/y.

Measured dose rates:

A series of measurements were taken to establish the *in situ* dose rates of the artificial background radiation and low radiation environments. The measurements were taken with a NaI detector (InSpector 1000 with an IRPOS-2: Stabilised 2" \times 2" probe), which gives real time values with a minimum dose rate equivalent of 10 nSv/h (0.088 mSv/y). Measurements of the unattenuated source yielded an absorbed dose rate of $D(0) = 29.9 \text{ mGy/y} \pm 0.5 \text{ mGy/y}$ and the 11-lead sheet-shielded dose rate of $D(3.3 \text{ cm lead}) = 1.067 \text{ mGy/y} \pm 0.2 \text{ mGy/y}$. The higher measured value than the theoretical value for the unattenuated source dose rate is consistent with measurement errors and backscattering of γ -rays from the walls of the lead castle. The measured dose rate within the castle with the lid closed and without the source was 0.0044 mGy/y, a reduction to $\sim 1\%$ natural background radiation. The measured local surface dose rates at Boulby ranged from 0.18 to 0.6 mGy/y. These were measured once at the beginning of the SELLR project with the same detector as stated above.

2.4. Intrinsic radiation from experimental apparatus

Due to the need to ensure an ultra low radiation environment, the internal intrinsic radiation of both the equipment and the samples had to be established to quantify the contribution to the final radiation dose. The radioactivity of the components used in this experiment was determined using γ -ray spectroscopy at the Boulby Underground Germanium Suite (BUGS). BUGS operates several low-background high purity germanium detectors in the Boulby Underground Laboratory (Scovell et al., 2018). This technique measures γ -ray emission associated with the decay of radioisotopes (usually U/Th/K along with anthropogenic radioisotopes) found in all materials and uses the detected flux to calculate a specific activity for each (Gilmore, 2011). This technique is widely used in material radio-assay for low-background particle physics experiments (Araújo et al., 2012; Abgrall et al., 2016; Aprile et al., 2017) where even small contaminations can severely impact detector sensitivity to rare

event searches. For SELLR, the plate readers, plates and 500 mL of bacterial growth media (with and without bacteria) were assayed. Each sample was placed on a germanium detector for approximately one week. From this, it was possible to determine that the potassium activity of all the constituent parts was substantially sub-dominant to the potassium activity of the bacteria. Table 2 shows the bacterial radiation contribution.

The highest internal nuclide activity from the bacterial sample was potassium (^{40}K), which undergoes β -decay (1311 keV end point energy with an 89% branching ratio) and electron capture (10.7% branching ratio, almost always followed by a 1461 keV γ -ray emitted from the ^{40}Ar daughter). Values of other nuclides are upper limits as they are below the sensitivity of the detector. The expected dose rate due to potassium is:

$$D_{K-40} = 0.0273 \text{ mGy/y} \pm 0.00352 \text{ mGy/y}$$

and the expected maximum absorbed dose rate due to other nuclides is given by:

$$D_{\text{other}} < 0.17104 \text{ mGy/y}$$

Although the expected dose rate of nuclides (excluding potassium) would exceed the levels necessary for the low radiation experiments, these values are the expected maximum calculated at 90% confidence level. The true values - and, by extension the true dose rate - are therefore highly likely to be below the maxima shown in table 2 and below the expected dose rate. To further constrain the actual experimental radiation environment, additional simulations and in situ measurements were also carried out (see sections 2.3 and 2.5).

2.5. Radiation simulation

To understand the radiation environment in the lead-lined **castle**, the entire experimental set up was constructed in GEANT4, and known radiation sources were added to simulate the resulting radiation. GEANT4 is a toolkit using Monte Carlo methods for simulating the passage of particles through matter (Agostinelli et al., 2003). This simulation was performed to provide insight into possible daughter particle radiation that must be taken into consideration when establishing the artificial background and low radiation environments. Radiation from a variety of sources was simulated, including the ^{137}Cs source, radon from the air, activities in the surrounding bedrock and lead in the **castle**. Uranium and thorium decay chains were assumed to be in secular equilibrium. It was concluded that γ -rays from the decay chains of uranium-thorium-potassium contamination in lead and the decay products of airborne radon could result in a ~ 0.1 mGy/y contribution to the radiation background in both halves of the **castle**. However, due to the higher dose contribution of the growth medium (see section 2.4) the contribution of the airborne radon was discounted. Additionally, the simulation showed the ^{137}Cs source used to produce the artificial radiation environment resulted in an additional dose rate of 314 nGy/year in the shielded, low radiation part of the **castle**. This value is however low enough to not significantly contribute to the low radiation environment.

2.6. Model organism selection

Bacillus subtilis (strain 168, DM 402) and *Escherichia coli* (MG 1655) were chosen as model organisms to test the effect of low radiation doses on microbial growth. Both microbes are well-

There's plenty of room at the bottom: Low radiation as a biological extreme

characterised environmental bacteria that can be cultured in the laboratory and were chosen to represent Gram positive and negative bacteria, respectively. Both have approximately the same, short doubling time of ~ 20 min, which allows for an efficient collection of multiple growth curves. Neither is classed as radio-resistant. We deliberately chose these characteristics to maximise the likelihood of detecting a response to the low radiation doses.

2.7. Cell growth set-up

B. subtilis and *E. coli* were grown in the artificial background and low radiation environments for 24 and 48 h. In some experiments, the dose rate was increased to $10 \times$ and $100 \times$ artificial surface background radiation by adjusting the distance of the ^{137}Cs source to the samples. The gradient of the bacterial exponential growth phase (growth rate) was determined, to establish whether either radiation environment had an effect on the bacterial growth. In addition to affecting growth rate, previous research (Satta et al., 1995) has claimed that prolonged exposure to a low radiation environment leaves cells more sensitive to stress factors, such as toxins or exposure to brief increases in radiation. To investigate this hypothesis, *B. subtilis* and *E. coli* were subjected to brief UVC radiation exposure after growth in the different radiation environments (see section 2.9).

2.8. Bacterial growth experiments

Because of the limited access to the mine, the growth experiments were performed in blocks of 5 - 6 days at a time. The terminology of “week 1, 2..” refers to the week in which the set of experiments was carried out, for ease of comparison (see table 3 for experimental nomenclature). To obtain aliquots of bacteria for the experiments, monocultures of *B. subtilis* and *E. coli* were grown at 37°C before being frozen into 25% glycerol 1 mL-aliquots and stored at -80°C at the University of Edinburgh. To obtain overnight cultures for experiments, 40 mL of nutrient broth was inoculated with 10 µL of thawed aliquots and cultured overnight at 37°C before use in the experiments. This was done at the overground laboratory at Boulby. The culture was then brought down to the underground laboratory at the beginning of each experimental run. At the beginning of each growth experiment, 5 µL of culture was added to 84 wells of a 96-well plate containing 200 µL of fresh nutrient broth. The last row of the plate contained only nutrient broth to serve as a blank for the plate reader. *B. subtilis* was grown at 30°C and *E. coli* at 37°C respectively for 24 h, with optical density (OD₆₀₀) measurements being taken every 20 minutes to coincide with both bacterial doubling times. For the 48 h experiments, 5 µL of the 24 h culture was added to 84 wells of a 96-well plate containing 200 µL of fresh nutrient broth and grown for 24 h under the same conditions as it was in the previous 24 h. The reason for having both 24 and 48 h growth data was to determine whether an extended period of exposure to the low radiation environment accentuated any observed responses in bacterial growth. This also gave us information on an increased number of generations, with 48 h resulting in ~ 144 generations for both *B. subtilis* and *E. coli*.

2.9. UV radiation exposure

In order to assess whether the microbes' ability to cope with stress was influenced by growth in the various radiation environments, cells were exposed to a short pulse of UVC and their viability was assessed by plating on Oxoid (LP0011) nutrient agar. Immediately after being grown for the specified amount of time in the respective radiation environments, the plates were irradiated with a

monochromatic UVC lamp ($\lambda = 254 \text{ nm}$; $I = 11.2 \text{ W/m}^2$; formally UVP Company now Analytik Jena AG) at distance of 5 cm for 30 s. Triplicates were selected from random wells of each condition and serially diluted by a factor of 10^6 . One hundred microlitres of each triplicate was plated in triplicate on nutrient agar and cultured overnight at 37°C . All steps of the UVC exposure and cultivation were conducted underground.

2.10. Experimental nomenclature

Three replicate low and artificial background radiation experiments were run with *B. subtilis*. For clarity when presenting these results, each set of experiments will be referred to with the nomenclature in Table 3 to differentiate between experimental runs. Whether the data is from the low or artificial background environment, and whether it is from the 24 or 48 h growth curves, will be stated in the text and figures. Henceforth, the one half of the experimental set up with 1% background radiation environment will be referred to as the “low” condition. The experimental set up involving enhanced radiation (background, $10 \times$ and $100 \times$ background radiation) containing the ^{137}Cs source will henceforth be referred to as the “artificial background” condition.

2.11. Analysis

All plate reader experiments were performed using 84 replicates; UV experiments were performed in triplicate with each triplicate plated on three agar plates. Numbers in figures show averages of replicates, error bars show standard error (s.e.) among replicates. Gradients of the exponential growth phases were established by an automated, linearly-fit trendline in excel. Error of gradients was calculated using the LINEST function in excel. Growth rates were calculated with the equation:

$$\mu = \frac{\ln OD_2 - \ln OD_1}{t_2 - t_1}$$

which when converted to the decadic logarithm becomes:

$$\mu = \frac{2.303(\log OD_2 - \log OD_1)}{t_2 - t_1}$$

OD_1 and OD_2 as well as respective t_1 and t_2 were determined in each growth curve as the beginning of the exponential growth phase. Statistical analysis was performed using one-way ANOVA and two-tailed unpaired equal variance Student's *t*-tests, where $p < 0.05$ was considered significant. Analysis of the bacterial growth curves focused on the linear exponential growth phases, which were most comparable and were used as a proxy for the relative fitness of each sample. When a non-linear “shoulder” in the exponential growth phase was observed, only data from the primary linear phase was used to calculate gradients.

3. Results

3.1. Overview of bacterial growth in low and artificial background radiation

There's plenty of room at the bottom: Low radiation as a biological extreme

Variations in growth were observed between experimental runs, but not between the differing radiation conditions.

All *B. subtilis* samples progressed through the lag, exponential and stationary growth phases. The upper cluster of final yield data in fig. 3 contained data points from the 48 h B1 data, 24 h & 48 h B2 data, as well as the B3 24 h data, whilst the lower cluster contained all B3 48 h data points. Each growth curve showed the presence of a “shoulder” in the exponential growth phase approximately between 480 and 600 minutes into growth after which all experimental runs, except the B3 48 h artificial background, progressed to a secondary exponential phase.

The stationary phase was reached with varying ODs from 1.5 to 1.8 with the exceptions of both B3 low and artificial background samples that reached between 1.2 and 1.5.

The last two hours of the plate reader data showed the onset of the death phase in all samples. However, in B3 48 h low and artificial background samples, the OD dropped more rapidly to below 1 and 1.2 respectively. These variations between samples is addressed in the discussion.

To verify whether this lack of effect of the different radiation environments on the exponential growth phase could be extended to other bacteria, experiments were replicated using *E. coli* (fig. 4). Once again, there was grouping according to experimental run (24 h and 48 h), not according to radiation environment, with the 24 h run having a shorter lag phase than that of the 48 h run. Similar to *B. subtilis*, “shoulders” appeared in the growth curves at ~ 200 minutes and at 300 - 400 minutes. However, unlike the wide variation of OD in the stationary and death phase of *B. subtilis*, the *E. coli* stationary phase OD values remained between 1 and 1.2. Additionally, there was no obvious grouping in the final yield between the low and artificial background samples, in contrast to the *B. subtilis* samples. The *E. coli* death phase was not captured during the aliquoted time of the experimental run.

3.2. Effects of radiation on growth gradients and rates

Figure 5(A & B) show the growth gradients and growth rates, respectively, of each experimental condition during the linear exponential growth phase. There was no significant ($p > 0.05$) difference between the low and artificial background radiation growth gradient at either time condition in the *B. subtilis* samples. However, the *E. coli* growth gradient showed a significant ($p < 0.05$) difference between the two time conditions in the artificial background radiation. The *E. coli* growth gradients for 24 h and 48 h in artificial background radiation were $0.0866 (\pm 4.33 \times 10^{-3})$ and $0.0693 (\pm 3.92 \times 10^{-3})$ respectively. The growth rates in fig. 5(B) were calculated for the average of all 84 samples in each condition and reflect the general trends shown by the growth gradients in fig. 5(A).

3.3. *B. subtilis* exposure to $10 \times$ and $100 \times$ background radiation

The *B. subtilis* growth curves displayed the same characteristics as the previous experiments, exhibiting a grouping in the exponential phase according to time condition, not radiation environment, in addition to a “shoulder” in the growth curve, and variation in OD during the stationary and death phases (fig. 6). Despite a non-significant ($p = 0.13$) difference in growth gradient (fig. 5(A)) and difference growth rate (fig. 5(B)) between the $10 \times$ and $100 \times$ artificial background radiation values, neither the $10 \times$ or $100 \times$ values were significantly ($p > 0.05$) different from the corresponding low radiation values in experiments run simultaneously as controls (fig. 5(A)). This implies that the difference in radiation dose rate was not the variable affecting growth.

3.4. Response to stress

The final preconditioning experiment was carried out to determine whether cells grown in a low radiation environment are more susceptible to short-term imposed stress compared to cells grown with the artificial background radiation exposure. *B. subtilis* and *E. coli* cultures that had been growing in low and artificial background radiation environments for 24 and 48 h respectively were exposed to a short burst of UVC radiation ($\lambda = 254$ nm) for 30 s and plated to assess their post-irradiation viability (fig. 7). There was no significant ($p > 0.05$) difference in viability observed after stress exposure between the cells grown in low or artificial background radiation, or between the two organisms.

4. Discussion

Scientific investigation of the lethal consequences caused by high radiation exposure for both prokaryotic and eukaryotic organisms continues to shape the understanding of habitability and the limits of life. Although there is an abundance of off-planet, high-radiation environments of interest (e.g. Cockell et al., 2000; Marion et al., 2003), naturally occurring low radiation environments represent equally important extreme conditions of potential habitability to examine. These might include subsurface environments with low abundances of radioactive minerals, older planets with decayed radioactive minerals, solar systems containing stars with a lower ionising radiation output and regions of the galaxy with a quieter cosmic ray background, compared to the Earth.

4.1. Effects of radiation on bacterial growth

We did not observe significant effects of low, artificial background, or heightened radiation environments (10 and $100 \times$ surface background levels) on the linear exponential growth phase gradients of *B. subtilis* and *E. coli* when compared to the respective controls. This contradicts the LNT model which predicts increasing levels of radiation would have an increasing effect on growth rate from a zero dose upwards. These data also contradict the hormesis model, that predicts a beneficial effect on growth rate at low levels of radiation. Instead, the data point to a threshold model for bacterial growth, whereby at low levels of radiation below background and up to 100 times background no effect is observable on growth. It should be noted that although the SELLR experimental set up was designed to observe the bacterial responses without bias towards any particular model, the set up may unintentionally favour certain models. We cannot rule out that hormesis takes place between any of the tested dose rates of $1 \times$, $10 \times$ and $100 \times$ background radiation. Future studies may wish to investigate a further narrowing of dose rates to more accurately rule out hormesis below our chosen radiation threshold.

We did observe variations in our experiments. The variation of lag phase is observed in all experiments performed. However, the variation in samples is grouped by experimental run and not by radiation environment, making the difference in radiation dose rate unlikely to be the cause. The variation in lag phase length is most likely due to small discrepancies in starting concentration of the samples. This experimental problem is not applicable to the exponential growth phase as it will be comparable regardless of small variations in starting inoculum.

An additional artefact in the *B. subtilis* and *E. coli* growth curves is the presence of a “shoulder” in the exponential growth phase. This may be a result of the bacteria exhausting the primary nutrient

There's plenty of room at the bottom: Low radiation as a biological extreme

source and switching to a secondary one, which is probable in the nutrient-rich media used. We did not use this secondary growth in our calculations.

The final growth yield data also show a wide diversity of OD values for *B. subtilis*. Factors that explain this may include, for example, cell lysis resulting in the release of various organics, which would influence cell growth and thus the final optical density measurement. Previous research has used final yield OD data as a measure of microbial responses to low radiation conditions (e.g. Castillo et al., 2015). However, the different contributions of cell growth, stationary phase and cell death will determine final OD readings and can therefore make the final OD interpretation potentially unreliable. As the exponential phase provides a measure of the rate of cell division and thus the effects of the radiation environment on cell reproduction, we consider this to be a good way to measure the effects of radiation.

Although the Boulby data challenges the LNT and hormesis models for low radiation doses, the indication that low dose rates do not impede bacterial growth is not unexpected. The empirical lethal dose necessary to sterilise 90% ("lethal dose" LD₉₀) of *E. coli* MG 1655 is 700 Gy (2.59×10^8 higher than the 24 h total dose received by Boulby control cells) (Daly et al., 2004). Although no LD₉₀ data could be found for vegetative *B. subtilis* cells, the majority of life is not radio-resistant and can be killed by < 500 Gy (Moeller et al., 2010; Slade and Radman, 2011). The cells subjected to 24 h of artificial background radiation in our experiment are exposed to a total dose of 2.7 µGy (or 5.4 µGy for 48 h experiments), whereas the cells' total dose after 24 h in the low radiation environment amounts to 0.012 µGy (0.024 µGy for 48 h experiments). Therefore, the total dose experienced by cells would have to be up to 8 orders of magnitude higher than the artificial background total dose to reach LD₉₀ (Daly, 2012). This also explains why, when the artificial background dose rate is increased by $10 \times$ and $100 \times$, it is unsurprising that no effect on bacterial growth was observed.

To further understand the scale at which radiation dose can influence cell viability, the likelihood of radiation-cell interaction must be considered. This likelihood was calculated by Lampe *et al.* (Lampe et al., 2016) for evolutionary research in the Modane Underground Laboratory (LSM) at 1.7 km below the Fréjus Peak (4.8 km water equivalent (Piquemal, 2012), Boulby is 3 km water equivalent (Robinson et al., 2003)). Using 7.4×10^{-4} mutations per generation as the spontaneous point mutation occurrence in *E. coli* (Barrick et al., 2009), they calculate that such natural mutation occurrences would happen at a 1.04×10^2 higher frequency than what they calculate for radiation-cell interactions. They compute that a dose rate of $> 20 \mu\text{Gy/h}$ ($= 175 \text{ mGy/y}$) is required for the radiation-cell interactions to have a detectable impact above the natural mutation "noise". These calculations further explain the lack of observable difference between the Boulby low, artificial background, and $100 \times$ artificial background (100 mGy/y) radiation dose rates on bacterial exponential growth. These data suggest that the radiation levels studied here are much lower than those required for significant cellular damage and reduction in growth.

Although the growth data may not rule out sublethal biochemistry and genetic responses caused by low radiation, as suggested by Castillo *et al.* (Castillo et al., 2015; Castillo et al., 2018), growth (i.e. reproduction) is the ultimate metric of whether an organism is able to expand into a habitat. If sublethal effects are present, they remain insufficient to ramify through metabolic pathways to impede growth rates. Based on our study, we can assume that exposure to an extremely low radiation environment would not negatively impact life.

4.2. Response to stress

Previous research has put forward the hypothesis that growth in a low radiation environment causes heightened sensitivity to stress factors. Satta *et al.* (Satta et al., 1995) demonstrated that *Saccharomyces cerevisiae* had increased sensitivity to the carcinogen methyl methanesulfonate after growth in a low radiation environment (0.6 $\mu\text{Gy/d}$, compared to Boulby's 0.012 $\mu\text{Gy/d}$). Additionally, it has been suggested (Castillo et al., 2015) that cultures grown in low radiation for 24 h are less able to respond to reactive oxygen species (ROS) attacking the cell. ROS can be produced by radiation interacting with water (Ward, 1988), with higher dose rates causing higher ROS production. It was proposed that the lack of radiation, and by extension lower ROS production, fails to “prime” cells’ response pathways for internal ROS removal, resulting in ROS accumulation and consequent stress.

This prompted our own experiment using UVC radiation exposure as a stress factor to determine whether there was any difference in cell sensitivity to stress after growth at low radiation versus growth under artificial surface radiation. The cellular response was measured in terms of cell viability after UV exposure. The results show no significant difference in either *B. subtilis* or *E. coli* viability grown at low or artificial background radiation dose rates.

These results suggest that even if there are biochemical effects in cells at below background radiation levels that are not manifested in growth, they also do not affect a cell’s short-term response to oxidative and radiation stress imposed by UV radiation.

4.3. Threshold model

Figure 8 synthesises the results obtained in this work. Between the total dose range of 10^{-8} - 10^{-4} Gy our data suggests a threshold model where there is no effect on bacterial growth at this dose and lower. The lowest total dose received by the Boulby samples was that of the 24 h exposure at the artificial background dose rate of 0.01 mGy/y resulting in a total dose of 2.74×10^{-8} Gy; the highest total dose received was the $100 \times$ the artificial background sample resulting in 2.74×10^{-4} Gy. Above this threshold but below doses that lead to loss of viability (10^{-4} - 10^2 Gy), we cannot rule out hypersensitivity or hermetic effects. There may be many additional “saddle points” in the model depending on factors such as dose-rate and organism-specific effects. Thus, the possibility of hypersensitivity or hormetic effect models may still be valid. Our results also focus on the exponential growth phase. However, if sublethal biochemical effects occur at below background radiation levels, as has been suggested for other cells (e.g. Kawanishi et al., 2012; Castillo et al., 2018), then it could suggest that different models apply depending on the scale or process considered and that no universal model for low radiation effects exists.

The data presented here indicate that there are no significant effects at either ultra low, artificial terrestrial surface, or heightened radiation dose rates on bacterial growth, and no difference in bacterial sensitivity to stress in the form of shortwave UV exposure after growth in low radiation environments. Thus, the preliminary data indicate that the extreme condition of low radiation in an environment would not restrict microbial expansion in a potential habitat. Additionally, the lack of a measurable linear effect or beneficial effects of low-level radiation suggest that both the LNT and hormesis models are inapplicable at these doses. Instead, a threshold model is suggested with no observable effect on viability below the terrestrial surface radiation dose up to a critical level exceeding the surface dose. The “critical threshold model” that emerges from our data might be

applicable to a wide range of physiological processes in prokaryotes and be a useful model to consider in future low radiation environment studies. However, the further questions raised by our data show that further research on life in low radiation environments is merited.

5. Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

6. Author Contributions

Internal dose measurements were performed by P.S., internal and external dose calculations were carried out by L.C., external dose measurements were carried out by S.P. and L.C., GEANT4 simulations were performed by A.N. under the supervision of A.StJ.M., growth measurements were performed by J.W. with the help of C.T. and E.M., the manuscript was written by J.W. with help from C.S.C., A.StJ.M. and P.S.

7. Funding

J.W. was funded by the UK Space Agency Aurora Studentship grant (STFC ST/M003612/1); C.S.C. was funded by the Science and Technology Facilities Council grant (ST/R000875/1); A.N. was funded by the Commonwealth Scholarship Commission grant (BDCS-2016-49); A.StJ.M. was funded by the Science and Technology Facilities Council grant (ST/N000269/1); L.C. was funded through the Trinity College MSc Physics programme; Boulby Underground Laboratory staff were funded by core Science and Technology Facilities Council facility funding.

8. Acknowledgments

We would like to acknowledge the technical assistance provided by Scott McLaughlin and general assistance provided by Barbara Suckling and Louise Yeoman.

9. References

Abgrall, N., Isaac J. Arnquist, F. T. Avignone Iii, Henning O. Back, Alexander S. Barabash, F. E. Bertrand, Melissa Boswell et al. "The MAJORANA DEMONSTRATOR radioassay program." *Nuclear Instruments and Methods in Physics Research Section A: Accelerators, Spectrometers, Detectors and Associated Equipment* 828 (2016): 22-36.

Agostinelli, Sea, John Allison, K. al Amako, John Apostolakis, H. Araujo, P. Arce, M. Asai et al. "GEANT4—a simulation toolkit." *Nuclear Instruments and Methods in Physics Research Section A: Accelerators, Spectrometers, Detectors and Associated Equipment* 506, no. 3 (2003): 250-303.

Aprile, Elena, J. Aalbers, F. Agostini, M. Alfonsi, F. D. Amaro, M. Anthony, B. Antunes et al. "The XENON1T dark matter experiment." *The European Physical Journal C* 77, no. 12 (2017): 881.

Araújo, H. M., D. Yu Akimov, E. J. Barnes, V. A. Belov, A. Bewick, A. A. Burenkov, V. Chepel et al. "Radioactivity backgrounds in ZEPLIN–III." *Astroparticle Physics* 35, no. 8 (2012): 495-502.

Barrick, Jeffrey E., Dong Su Yu, Sung Ho Yoon, Haeyoung Jeong, Tae Kwang Oh, Dominique Schneider, Richard E. Lenski, and Jihyun F. Kim. "Genome evolution and adaptation in a long-term experiment with *Escherichia coli*." *Nature* 461, no. 7268 (2009): 1243-1247.

Calabrese, Edward J., and Linda A. Baldwin. "Hormesis: the dose-response revolution." *Annual Review of Pharmacology and Toxicology* 43, no. 1 (2003): 175-197.

Castillo, Hugo, Donald Schoderbek, Santosh Dulal, Gabriela Escobar, Jeffrey Wood, Roger Nelson, and Geoffrey Smith. "Stress induction in the bacteria *Shewanella oneidensis* and *Deinococcus radiodurans* in response to below-background ionizing radiation." *International Journal of Radiation Biology* 91, no. 9 (2015): 749-756.

Castillo, Hugo, Xiaoping Li, Faye Schilkey, and Geoffrey B. Smith. "Transcriptome analysis reveals a stress response of *Shewanella oneidensis* deprived of background levels of ionizing radiation." *PloS One* 13, no. 5 (2018).

Chang, J. Ch, Susan F. Ossoff, David C. Lobe, Mark H. Dorfman, Constance M. Dumais, Robert G. Qualls, and J. Donald Johnson. "UV inactivation of pathogenic and indicator microorganisms." *Appl. Environ. Microbiol.* 49, no. 6 (1985): 1361-1365.

Cockell, Charles S., David C. Catling, Wanda L. Davis, Kelly Snook, Ray L. Kepner, Pascal Lee, and Christopher P. McKay. "The ultraviolet environment of Mars: biological implications past, present, and future." *Icarus* 146, no. 2 (2000): 343-359.

Costantini, David, and Benny Borremans. "The linear no-threshold model is less realistic than threshold or hormesis-based models: an evolutionary perspective." *Chemico-Biological Interactions* 301 (2019): 26-33.

Daly, Michael J., E. K[†] Gaidamakova, V. Y. Matrosova, A. Vasilenko, M. Zhai, Amudhan Venkateswaran, M. Hess et al. "Accumulation of Mn (II) in *Deinococcus radiodurans* facilitates gamma-radiation resistance." *Science* 306, no. 5698 (2004): 1025-1028.

Daly, Michael J. "Death by protein damage in irradiated cells." *DNA Repair* 11, no. 1 (2012): 12-21.

de Gonzalez, Amy Berrington, and Sarah Darby. "Risk of cancer from diagnostic X-rays: estimates for the UK and 14 other countries." *The Lancet* 363, no. 9406 (2004): 345-351.

de Vera, Jean-Pierre, Mashal Alawi, Theresa Backhaus, Mickael Baqué, Daniela Billi, Ute Böttger, Thomas Berger et al. "Limits of life and the habitability of Mars: the ESA space experiment BIOMEX on the ISS." *Astrobiology* 19, no. 2 (2019): 145-157.

Feinendegen, Ludwig E., Myron Pollycove, and Ronald D. Neumann. "Whole-body responses to low-level radiation exposure: new concepts in mammalian radiobiology." *Experimental Hematology* 35, no. 4 (2007): 37-46.

There's plenty of room at the bottom: Low radiation as a biological extreme

Feynman, Richard P. "There's plenty of room at the bottom." *California Institute of Technology, Engineering and Science Magazine* (1960).

Finch, Stuart C. "Radiation-induced leukemia: lessons from history." *Best Practice & Research Clinical Haematology* 20, no. 1 (2007): 109-118.

Ghiassi-Nejad, M., S. M. J. Mortazavi, J. R. Cameron, A. Niroomand-Rad, and P. A. Karam. "Very high background radiation areas of Ramsar, Iran: preliminary biological studies." *Health Physics* 82, no. 1 (2002): 87-93.

Gilmore, Gordon. *Practical Gamma-Ray Spectroscopy*. John Wiley & Sons, 2011.

Hamilton, E. I. "Terrestrial radiation—an overview." *International Journal of Radiation Applications and Instrumentation. Part C. Radiation Physics and Chemistry* 34, no. 2 (1989): 195-212.

Horneck, Gerda. "Radiobiological experiments in space: a review." *International Journal of Radiation Applications and Instrumentation. Part D. Nuclear Tracks and Radiation Measurements* 20, no. 1 (1992): 185-205.

International Agency for Research on Cancer. "Monograph on the evaluation of carcinogenic risks to humans: solar and ultraviolet radiation." *WHO* 55 (1992): 113-122.

Kawanishi, Masanobu, Katsuyuki Okuyama, Kazunori Shiraishi, Yatsuka Matsuda, Ryoichi Taniguchi, Nobuyuki Shiomi, Morio Yonezawa, and Takashi Yagi. "Growth retardation of *paramecium* and mouse cells by shielding them from background radiation." *Journal of Radiation Research* 53, no. 3 (2012): 404-410.

Lampe, Nathanael, David G. Biron, Jeremy MC Brown, Sébastien Incerti, Pierre Marin, Lydia Maigne, David Sarramia, Hervé Seznec, and Vincent Breton. "Simulating the impact of the natural radiation background on bacterial systems: Implications for very low radiation biological experiments." *PloS One* 11, no. 11 (2016).

Lorenz, Egon. "Radioactivity and lung cancer; a critical review of lung cancer in the miners of Schneeberg and Joachimsthal." *Journal of the National Cancer Institute* 5, no. 1 (1944): 1-15.

Lorenz, Egon. "Some biologic effects of long continued irradiation." *The American Journal of Roentgenology and Radium Therapy* 63, no. 2 (1950): 176-185.

Mancuso, M., E. Pasquali, P. Giardullo, S. Leonardi, M. Tanori, V. Di Majo, S. Pazzaglia, and A. Saran. "The radiation bystander effect and its potential implications for human health." *Current Molecular Medicine* 12, no. 5 (2012): 613-624.

There's plenty of room at the bottom: Low radiation as a biological extreme

Marion, Giles M., Christian H. Fritsen, Hajo Eicken, and Meredith C. Payne. "The search for life on Europa: limiting environmental factors, potential habitats, and Earth analogues." *Astrobiology* 3, no. 4 (2003): 785-811.

Mine, M., Y. Okumura, M. Ichimaru, T. Nakamura, and S. Kondo. "Apparently beneficial effect of low to intermediate doses of A-bomb radiation on human lifespan." *International Journal of Radiation Biology* 58, no. 6 (1990): 1035-1043.

Moeller, Ralf, Thierry Douki, Petra Rettberg, Günther Reitz, Jean Cadet, Wayne L. Nicholson, and Gerda Horneck. "Genomic bipyrimidine nucleotide frequency and microbial reactions to germicidal UV radiation." *Archives of Microbiology* 192, no. 7 (2010): 521-529.

Moeller, Ralf, Marina Raguse, Stefan Leuko, Thomas Berger, Christine Elisabeth Hellweg, Akira Fujimori, Ryuichi Okayasu, Gerda Horneck, and STARLIFE Research Group. "STARLIFE—An international campaign to study the role of galactic cosmic radiation in astrobiological model systems." *Astrobiology* 17, no. 2 (2017): 101-109.

Muller, Hermann J. "Nobel Prize Lecture." *Stockholm, Sweden, December 12 (1946)*.

Muller, Hermann J. "Damage to posterity caused by irradiation of the gonads." *American Journal of Obstetrics & Gynecology* 67, no. 3 (1954): 467-483.

National Research Council. *Health risks from exposure to low levels of ionizing radiation: BEIR VII phase 2*. Vol. 7. National Academies Press, 2006.

Olson, Axel R., and Gilbert N. Lewis. "Natural reactivity and the origin of species." *Nature* 121, no. 3052 (1928): 673-674.

Piquemal, F. "Modane underground laboratory: Status and project." *The European Physical Journal Plus* 127, no. 9 (2012): 110.

Preston, Dale L., Donald A. Pierce, Yukiko Shimizu, Harry M. Cullings, Shoichiro Fujita, Sachiyo Funamoto, and Kazunori Kodama. "Effect of recent changes in atomic bomb survivor dosimetry on cancer mortality risk estimates." *Radiation Research* 162, no. 4 (2004): 377-389.

Prise, K. M., M. Pinto, H. C. Newman, and B. D. Michael. "A review of studies of ionizing radiation-induced double-strand break clustering." *Radiation Research* 156, no. 5 (2001): 572-576.

Reichhart, Lindote, A. Lindote, D. Yu Akimov, H. M. Araújo, E. J. Barnes, V. A. Belov, A. Bewick et al. "Measurement and simulation of the muon-induced neutron yield in lead." *Astroparticle Physics* 47 (2013): 67-76.

Robinson, M., V. A. Kudryavtsev, R. Lüscher, J. E. McMillan, P. K. Lightfoot, N. J. C. Spooner, N. J. T. Smith, and I. Liubarsky. "Measurements of muon flux at 1070m vertical depth in the Boulby

There's plenty of room at the bottom: Low radiation as a biological extreme

underground laboratory." *Nuclear Instruments and Methods in Physics Research Section A: Accelerators, Spectrometers, Detectors and Associated Equipment* 511, no. 3 (2003): 347-353.

Satta, L., G. Augusti-Tocco, R. Ceccarelli, A. Esposito, M. Fiore, P. Paggi, I. Poggesi, R. Ricordy, G. Scarsella, and E. Cundari. "Low environmental radiation background impairs biological defence of the yeast *Saccharomyces cerevisiae* to chemical radiomimetic agents." *Mutation Research Letters* 347, no. 3-4 (1995): 129-133.

Scovell, P. R., E. Meehan, H. M. Araújo, J. Dobson, C. Ghag, H. Kraus, V. A. Kudryavtsev et al. "Low-background gamma spectroscopy at the Boulby Underground Laboratory." *Astroparticle Physics* 97 (2018): 160-173.

Shahbazi-Gahrouei, Daryoush, Mehrdad Gholami, and Samaneh Setayandeh. "A review on natural background radiation." *Advanced Biomedical Research* 2 (2013).

Shamoun, Dima Yazji. "Linear No-Threshold model and standards for protection against radiation." *Regulatory Toxicology and Pharmacology* 77 (2016): 49-53.

Shibamoto, Yuta, and Hironobu Nakamura. "Overview of biological, epidemiological, and clinical evidence of radiation hormesis." *International Journal of Molecular Sciences* 19, no. 8 (2018): 2387.

Siasou, Eleni, David Johnson, and Neil J. Willey. "An extended dose–response model for microbial responses to ionizing radiation." *Frontiers in Environmental Science* 5 (2017): 6.

Slade, Dea, and Miroslav Radman. "Oxidative stress resistance in *Deinococcus radiodurans*." *Microbiol. Mol. Biol. Rev.* 75, no. 1 (2011): 133-191.

Smith, Geoffrey Battle, Yair Grof, Adrienne Navarrette, and Raymond A. Guilmette. "Exploring biological effects of low level radiation from the other side of background." *Health Physics* 100, no. 3 (2011): 263-265.

Stebbing, A. R. D. "Hormesis—the stimulation of growth by low levels of inhibitors." *Science of the Total Environment* 22, no. 3 (1982): 213-234.

United Nations. Scientific Committee on the Effects of Atomic Radiation. *Report of the United Nations Scientific Committee on the Effects of Atomic Radiation: Fifty-sixth Session (10-18 July 2008)*. No. 46. United Nations Publications, 2008.

Wanebo, C. K., K. G. Johnson, K. Sato, and T. W. Thorslund. "Breast cancer after exposure to the atomic bombings of Hiroshima and Nagasaki." *New England Journal of Medicine* 279, no. 13 (1968): 667-671.

Ward, John F. "DNA damage produced by ionizing radiation in mammalian cells: identities, mechanisms of formation, and reparability." In *Progress in Nucleic Acid Research and Molecular Biology*, vol. 35, pp. 95-125. Academic Press, 1988.

10. Tables

Table 1: Potential problems facing low radiation experiments and the respective adjustments made for the SELLR experimental set up.

Potential issues for low radiation experimental set ups	SELLR Set Up adjustments
Lack of controls in identical conditions. In the case of underground experiments and ground controls, differences such as air pressure and humidity between surface and sub-surface environments can skew bacterial growth results.	Controls were situated underground with the low radiation samples. An artificial radiation source was used to provide surface and above surface radiation dose exposure; all other parameters were identical.
Model organisms used were known to be radio-resistant and would not show representative effects of low radiation exposure of the majority of microbes.	Well-characterised environmental model organisms were used to represent Gram positive & negative microbes without specific radio-resistance.
Use of end-point analysis of growth curves, which limit analysis of the full growth curve and the possible effects of below background radiation on different parts of the growth curve.	Analysis of radiation impact on the whole growth curve so that the linear exponential growth phase (growth rate) could be determined as well as final growth parameters.
Lack of replicates and sufficient time points to examine growth data.	Continuous measurement of growth curves to determine bacterial growth rate, large number of replicates, and experiment repetition.
Only two doses tested (background radiation & low radiation); insufficient to plot or rule out different models.	Low, artificial background radiation, 10× and 100× artificial background radiation doses were used in identical experimental set-ups for multiple dose data points.
Reliant on simulation of low radiation environment dose, with no <i>in situ</i> confirmation of real dose environment.	Simulation and <i>in situ</i> measurement of low and artificial background radiation environment.

There's plenty of room at the bottom: Low radiation as a biological extreme

Table 2: Dose contribution of nuclide series in the bacterial sample (growth media + *B. subtilis* inoculate)

Nuclide series	Activity in [Bq/kg]
$^{238}\text{U}(\text{e})$	-
$^{238}\text{U}(\text{l})$	<0.4
$^{232}\text{Th}(\text{e})$	<0.6
$^{232}\text{Th}(\text{l})$	<0.4
^{40}K	11.5 ± 1.5
^{60}Co	<0.1
^{235}U	<0.8

Table 3. SELLR experiment nomenclature for *B. subtilis* replicate experiments and *E. coli* experiments indicating growth time, radiation exposure and the week of experimental execution; art. = artificial.

Sample name	Growth time	Radiation exposure	Organism
B1	48 h	Low & artificial background	<i>B. subtilis</i>
B2	24 & 48 h	Low & artificial background	<i>B. subtilis</i>
B3	24 & 48 h	Low & artificial background	<i>B. subtilis</i>
10 ×	24 h	Low & 10 × background	<i>B. subtilis</i>
100 ×	24 h	Low & 100 × background	<i>B. subtilis</i>
<i>E. coli</i>	24 & 48 h	Low & artificial background	<i>E. coli</i>

11. Figures

Figure 1: Proposed models for experimental and theoretical correlations of radiation dose and the risk of damage to organisms, including the widely accepted LNT model. Models are based on high-dose data, extrapolating responses for lower dose ranges. High doses are defined as those causing observable, serious or irreparable damage to an organism. The dotted lines indicate the potential models that may be applicable to prokaryotes in the intermediate and low radiation dose ranges, i.e. either side of background radiation up until high doses. LNT = linear relationship between dose and risk of damage to an organism; sensitivity = a heightened risk of damage at lower doses; hormesis = high dose inhibition with low dose stimulation; threshold = no/negligible risk of damage below a specific dose.

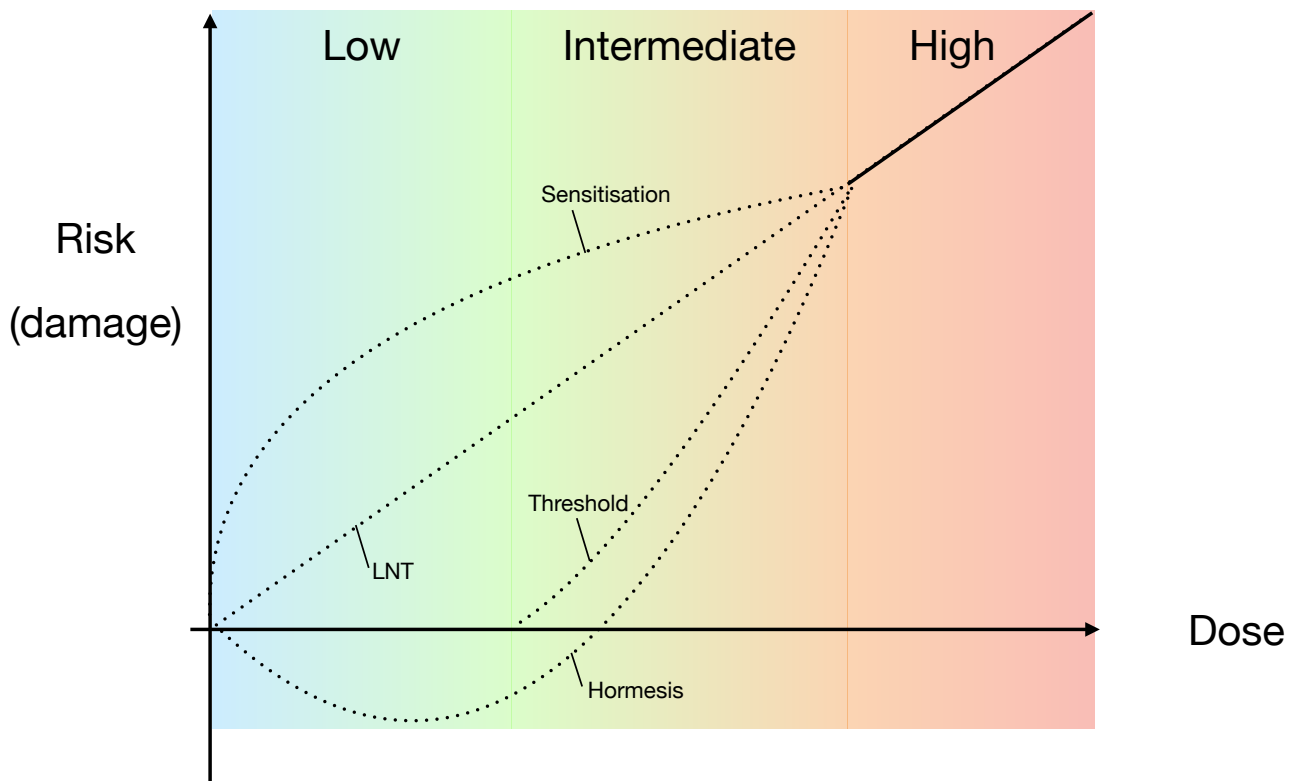
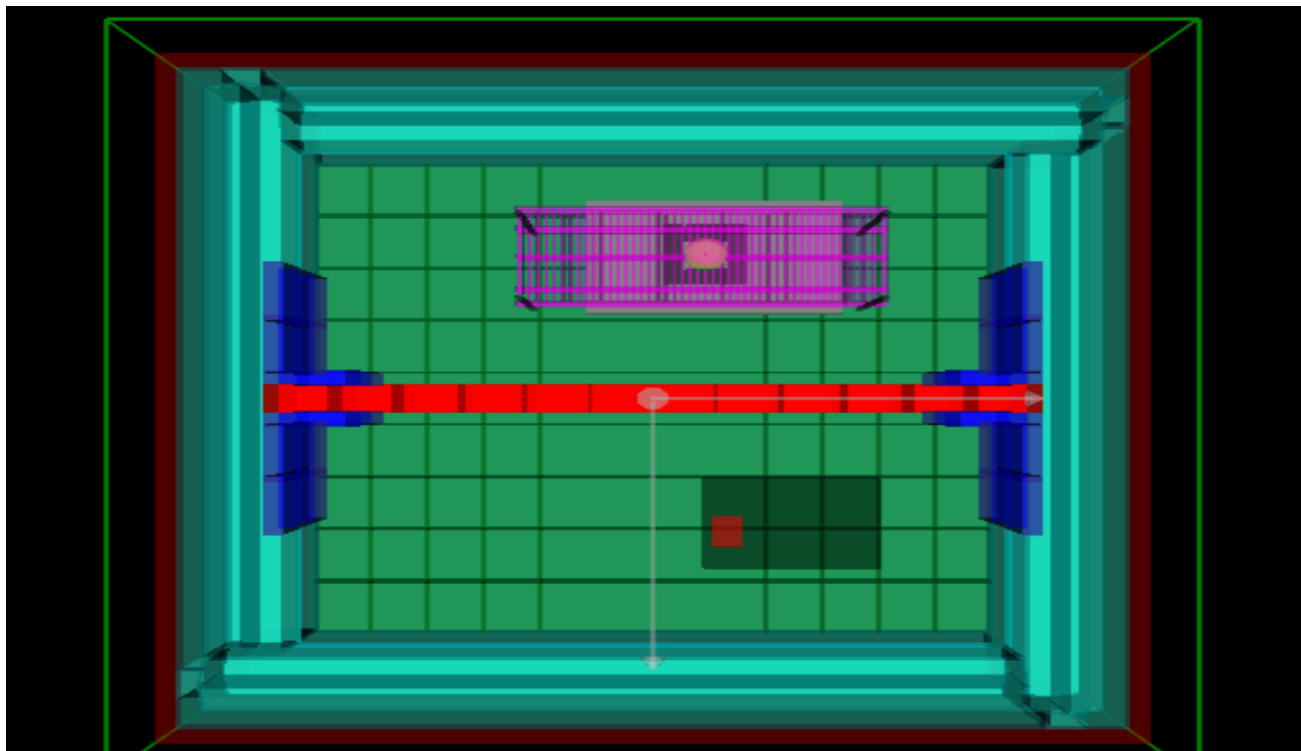


Figure 2: Geometry of lead-lined chamber in GEANT4, view from above. The lead wall is marked in red, separating the container into a low and artificial background radiation environment. The ^{137}Cs γ -source is located on the purple platform above the one plate reader, providing the artificial background radiation.



There's plenty of room at the bottom: Low radiation as a biological extreme

Figure 3: Average ($n = 84$) of *B. subtilis* growth curves at the determined low radiation (low) and artificial background radiation (“sim. background”) dose rate: 24 and 48 h denote time in hours that bacteria batch has spent in respective conditions; B1-3 = experiment run number. Error bars show \pm standard error ($n = 84$).

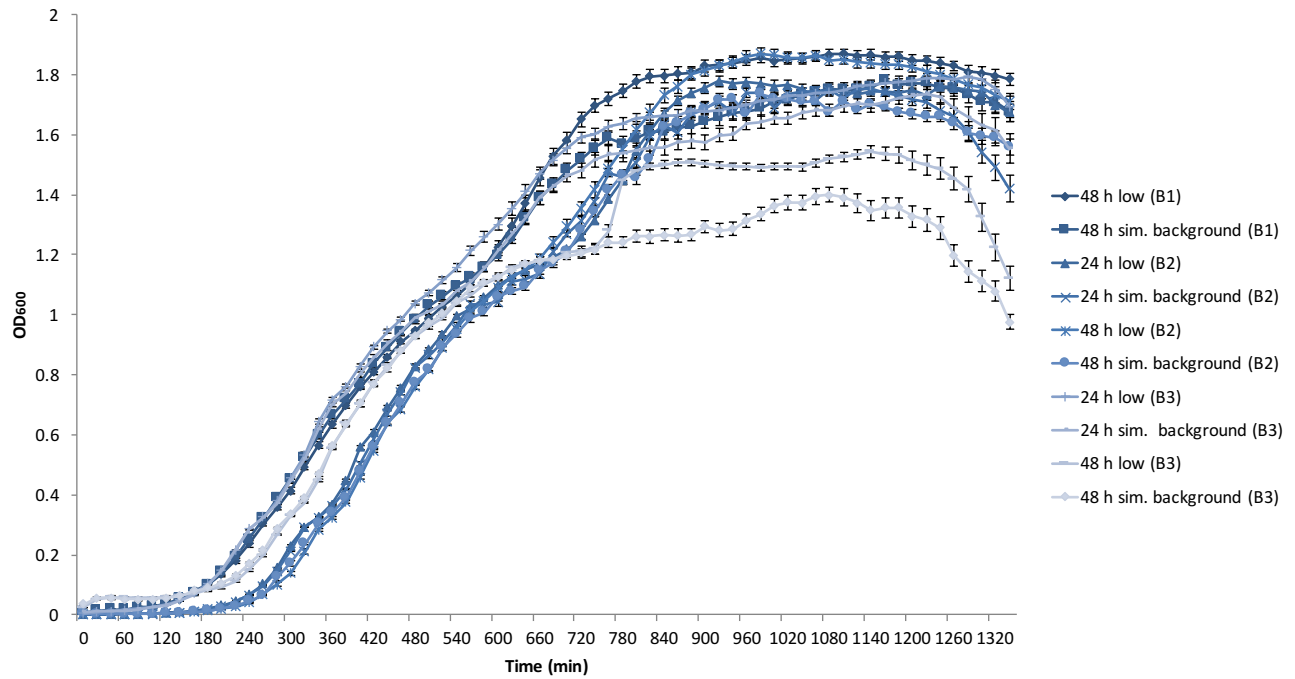


Figure 4: Average ($n = 84$) of *E. coli* growth curves at the determined low radiation (“low”) and artificial background radiation (“sim. background”) dose rate: 24 and 48 h denote time in hours that bacteria batch has spent in respective conditions. Error bars show \pm standard error ($n = 84$).

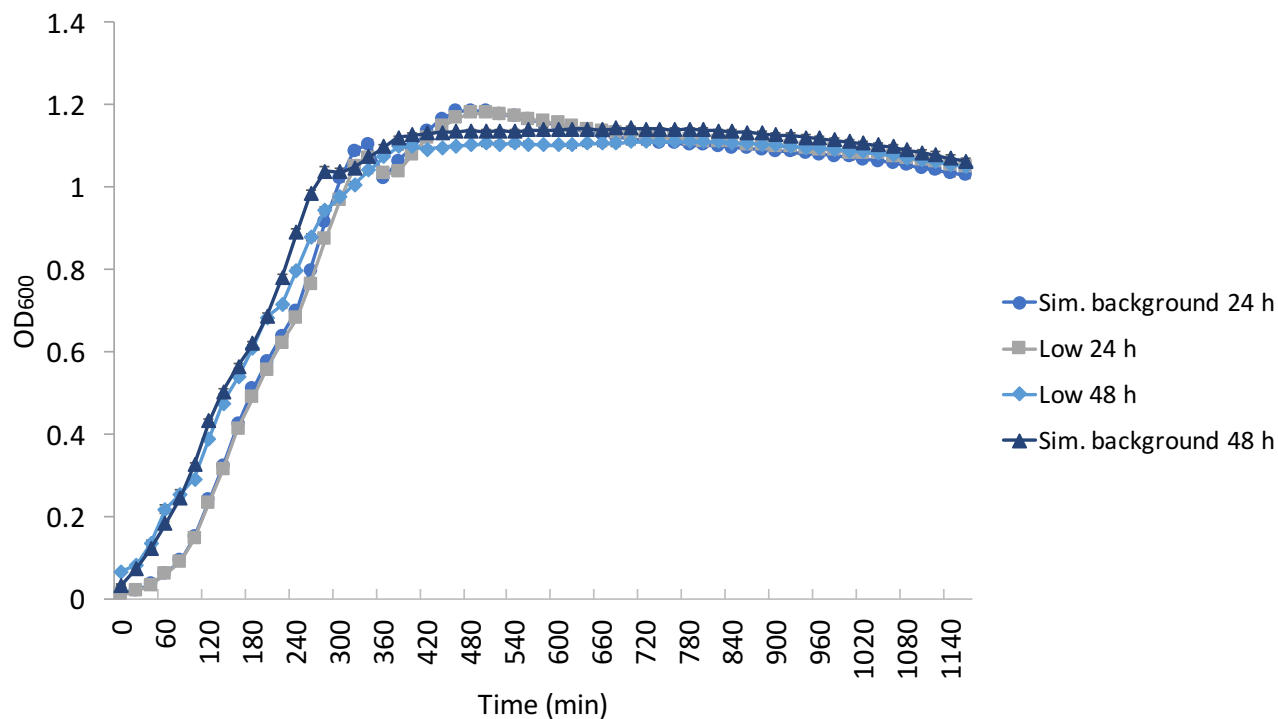
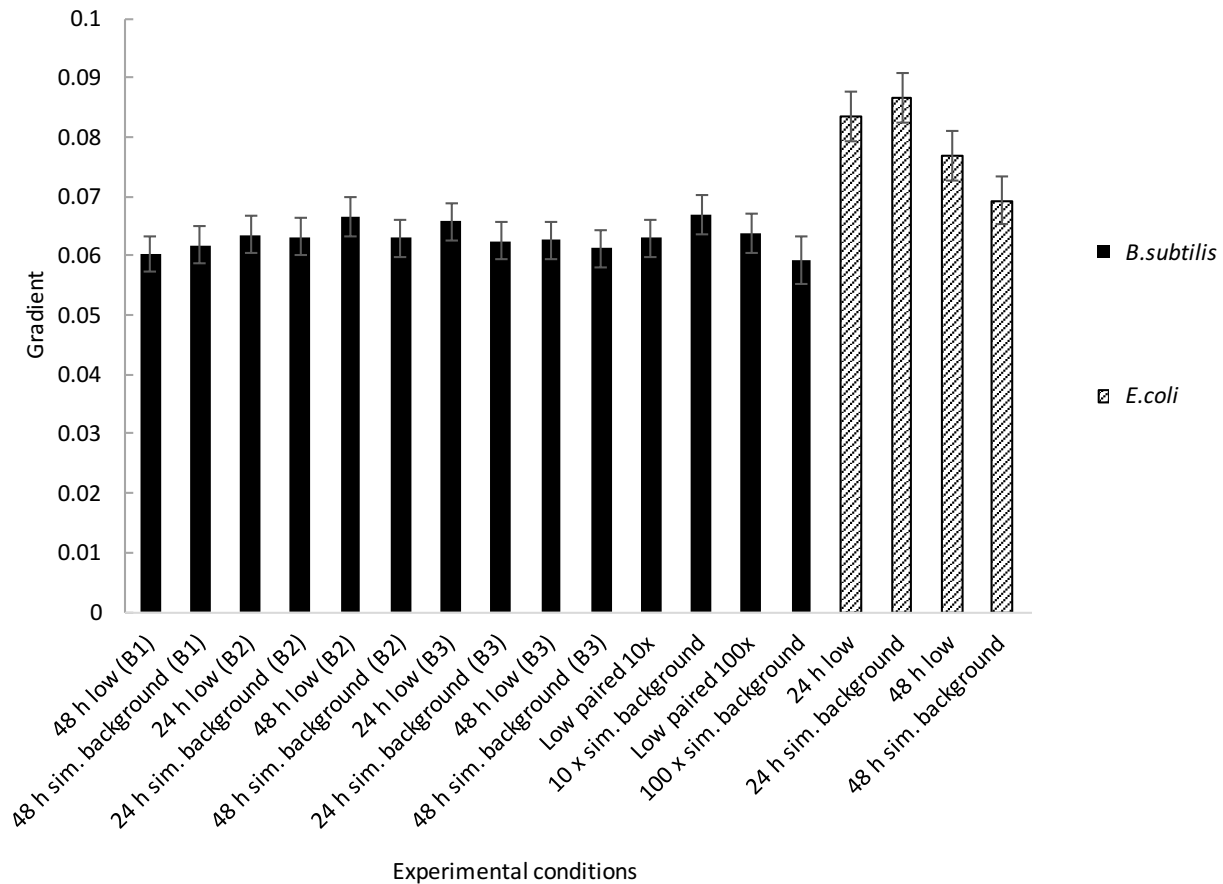
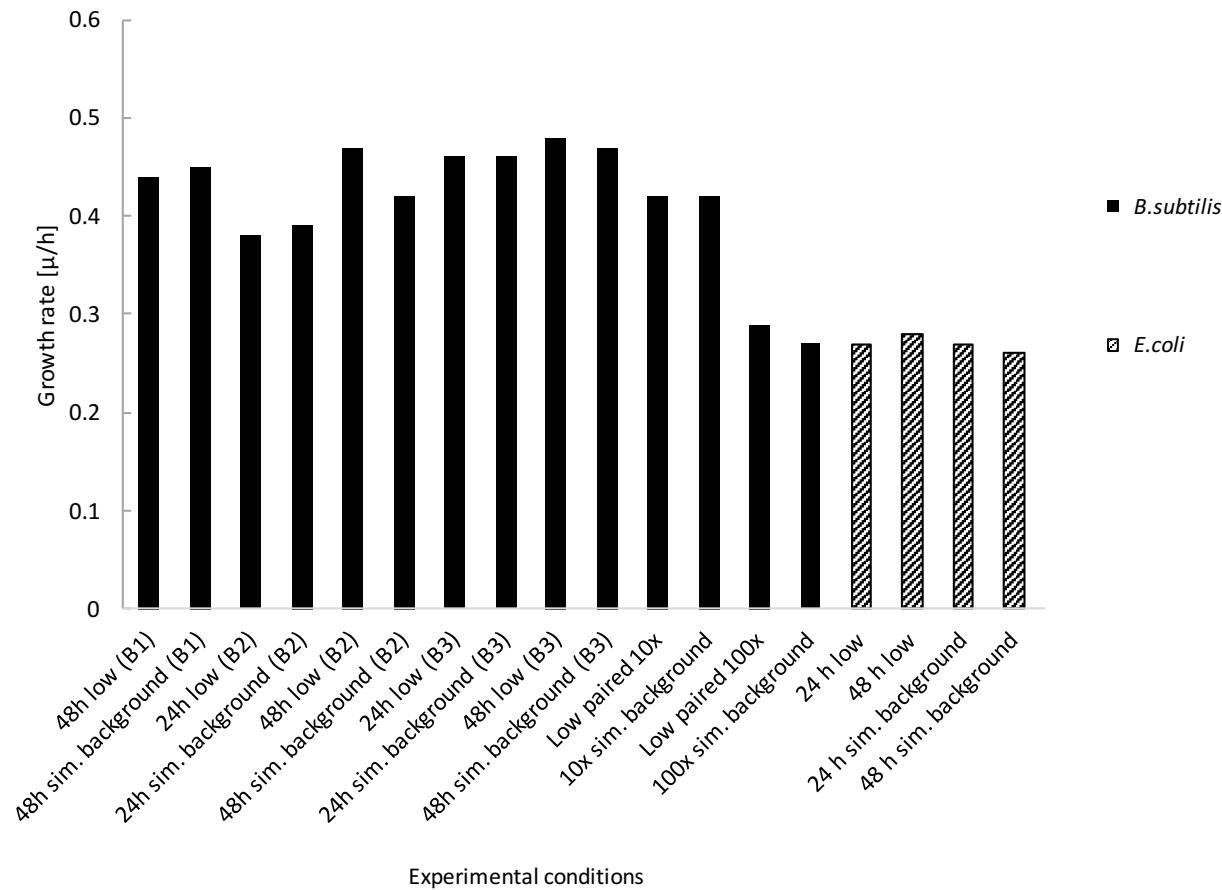


Figure 5: Growth gradients and rates of *B. subtilis* and *E. coli* linear exponential growth phases at the determined low radiation (“low”) and artificial background radiation (“sim. background”) dose rate: 24 and 48 h denote time in hours that bacteria batch has spent in respective conditions; B1-3 denote week number when the multiple *B. subtilis* samples were run; $10 \times = 10 \times$ the artificial background dose rate; $100 \times = 100 \times$ the artificial background dose rate; Low paired = low dose rate samples run parallel to $10 \times$ and $100 \times$ dose rates. Error bars show \pm error of gradient. “B.s” and “E.c.” stand for *B. subtilis* and *E. coli*, respectively.

(A): Growth gradients of *B. subtilis* and *E. coli* from the linear exponential growth phase.



(B): Growth rates of *B. subtilis* and *E. coli* from the linear exponential growth phase.



There's plenty of room at the bottom: Low radiation as a biological extreme

Figure 6: Average ($n = 84$) of *B. subtilis* growth curves at increased radiation dose rates: $10\times = 10\times$ the artificial background dose rate; $100\times = 100\times$ the artificial background dose rate; Low paired = samples run parallel at the low dose rate. Error bars show \pm standard error ($n = 84$)

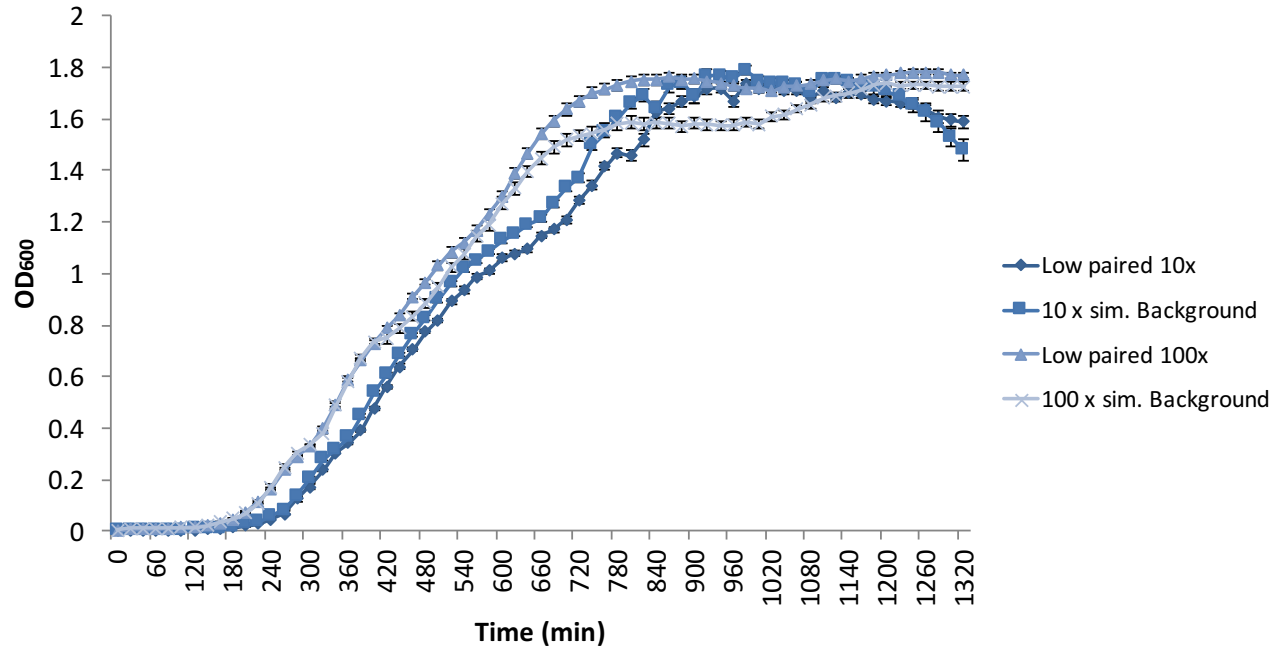
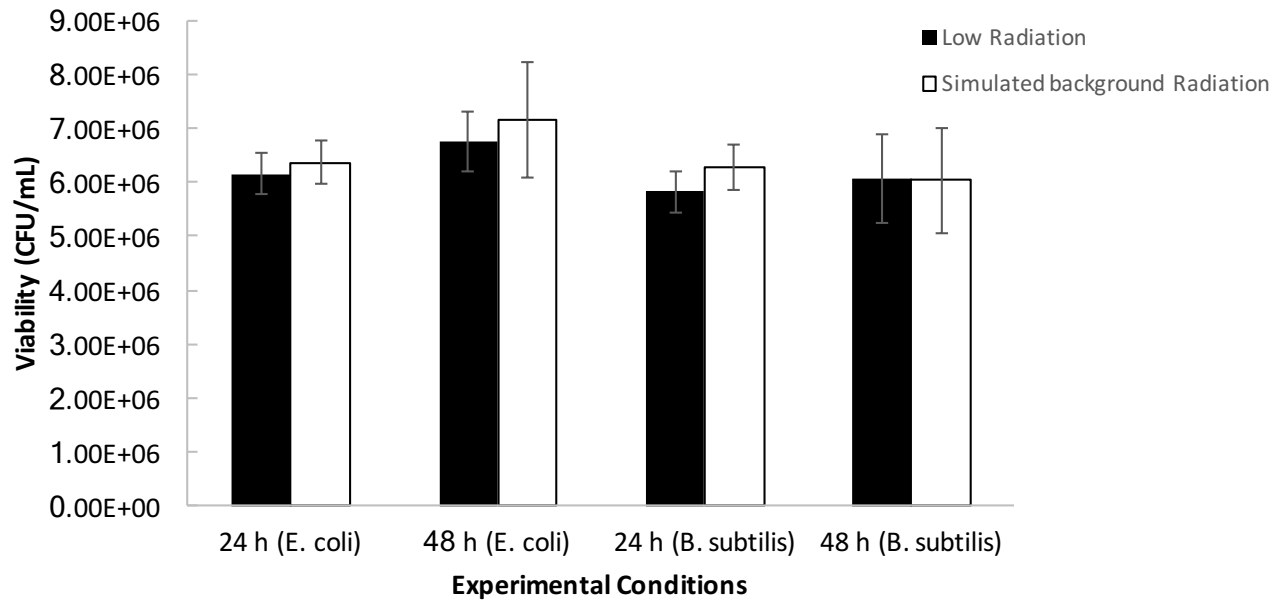


Figure 7: Averages ($n = 9$ per condition) of post-UV irradiation cell counts of *B. subtilis* and *E. coli* grown under the determined low radiation (“low”) and artificial background radiation (“sim. background”) dose rate: 24 and 48 h denote time in hours that bacteria batch has spent in respective conditions. Error bars show \pm standard error ($n = 9$ per condition).



There's plenty of room at the bottom: Low radiation as a biological extreme

Figure 8: Updated prokaryotic dose-response model with a linear relationship for higher doses & added SELLR (Subsurface Experiment of Life in Low Radiation) observations from sub-background to $100 \times$ background, representing the “low” dose range where we observe no difference in growth. Black dotted lines indicate hypothetical risk of damage at intermediate radiation exposure, which requires further research.

